TOXIC AND LATENT EFFECT OF THREE ISOLATED BOTANICAL CHEMICALS ON THE EGYPTIAN COTTON LEAFWORM, *SPODOPTERA LITTORALIS* (BOISD). NOCTUIDAE: LEPIDOPTERA

ELHAM F. M. ABDEL-RAHIM, M. E. M. AHMED, A. A. AHMED AND T. A. MOHAMED

1. Sils Agric. Res. Station, Plant Protection Res. Institute, ARC, Giza
2. Faculty of Science, Chemical division, El-Minia University

(Manuscript received 27 May 2007)

### Abstract

The toxic effect of three chemicals were isolated from plants *Anaphalis margaritacea*, *Chenopodium ambrosioides* and *Ambrosia maritima* against the second and fourth instar larvae of the cotton leafworm, *Spodoptera littoralis* was bioassayed in the laboratory tests. The molecular formula of these chemicals were C_{10}H_{10}O_{2}, C_{12}H_{20}O_{4} and C_{12}H_{24}O_{3}, respectively. The 2nd and 4th instar larvae were left to contact for 24h with the residual film of each chemical at different concentrations. The first chemical (C_{10}H_{10}O_{2}) was the most toxic against the two instar larvae with LC_{50} values of 6 and 28 P.P.M. for both instars, while the third chemical was the least toxic one with LC_{50} values of 60 and 94 P.P.M. for the two instar larvae, respectively. The biological parameters of the treated larval instars were obviously affected significantly with the first chemical. Therefore, the larval and pupal duration were prolonged, the pupation and adult emergence percentages were decreased, the pupal weight was reduced. Also, the fecundity and fertility were retard and the longevity was shorted, also the sex ratio of the emerged adults was shifted in respect to control. The Pupal and adult malformation percentages were recorded as a result of the larvae treated of 4th instar with these chemicals at the LC_{50} values. Most of the tested biological activities of the treated larvae with the three botanical chemicals were more affected with the third chemical (C_{12}H_{20}O_{4}) compared with the second chemical (C_{12}H_{24}O_{3}) treatments than the second chemical (C_{10}H_{10}O_{2}) treatments.

### INTRODUCTION

The cotton leafworm, *Spodoptera littoralis* (Boisd) is one of the major pests that cause a considerable damage to many of the important vegetables and field crops in Egypt. The massive application of pesticides has created serious problems such as the build-up of pest resistance, the upsetting of natural balance, acute and chronic hazards to man and animals. It is therefore necessary to complement our reliance on synthetic pesticides with less hazardous, safe, and biodegradable substitutes. The use of natural products of plant origin is a new trend that preserve the environment from contamination with harmful toxicants. The most familiar one of these chemicals in the
recent years is azadirachtin that isolated from the fruits of Melia azedarach and seeds of Azadirachta indica (Meliaceae). Azadirachtin exhibits extremely low acute mammalian toxicity, yet it is very effective as control agent for many insect groups (Sharma et al., 1992). Besides azadirachtin, other plant derived extracts and phytochemicals have been shown to possess insecticidal activity and some morphogenetic effects. The insecticidal activity of several isolated phytochemicals was evaluated against several Lepidopteous insects particularly S. littoralis (Simmonds et al., 1990; Amr et al., 1995). Also some morphogenetic effects of some plant derived chemicals against several Lepidoptera species were recorded (Urinova et al., 1989; Abo-El-Ghar et al., 1994).

The present study aimed to evaluate the insecticidal and morphogenetic activity of some plant derived chemicals against S. littoralis.

**MATERIALS AND METHODS**

1. **Insect rearing**

   The cotton leaf worm, S. littoralis was reared in the laboratory for several generations at room temp. ranged between 25 - 28°C and 60 - 65% R.H. The instar larvae were fed on castor bean leaves, *Ricinus communis* (L.) in a wide glass jars until pupation period and adults emergence. The newly emerged adults were mated inside glass jars supplied with a piece of cotton wetted with 10% sugar solution as feeding source for the emerged moths and branches of Tafla leaf (*Nerium oleander* L.) or castor bean leaves as an oviposition site (El-Defrawi et al., 1964). Egg masses were kept in plastic jars until hatching.

2. **Material used**

   Three chemicals were isolated from three plants: The first plant, *Anaphalis margaritacea* (Asteraceae). It was a stoloniferous plant, the common name is Pearly everlasting, the samples of this plant collected from North American. The second plant, *Chenopodium ambrosioides* (Chenopodiaceae), and the third plant, *Ambrosia maritima* (Compositae), the samples of the latter plants were collected from North Sinai. All samples were collected and separated by Prof. Dr. Ahmed A. Ahmed in Biochemical Division, Faculty of Science, El- Minia University. The molecular formula for these botanical chemicals were: \(C_{13}H_{22}O_8\), \(C_{27}H_{52}O_{14}\) and \(C_{16}H_{16}O_7\), respectively.
The chemical structure of the compound as following:

- **The first chemical**:
  - Molecular formula: C_{19}H_{24}O_4
  - Molecular weight: 256 mg
  - Systematic name: dihydroxy dibenzoyl acid (phenolis)

![Structure formula of the first chemical]

- **The second chemical**:
  - Molecular formula: C_{19}H_{20}O_4
  - Molecular weight: 578 mg
  - Systematic name: chrysophan (glucoside)
  - Chemical name: K-3,7-dihydro- Oside

- **The third chemical**:
  - Molecular formula: C_{19}H_{18}O_7
  - Systematic name: 5,6 dihydroxy-3,4,7 trimethoxy flavone
  - Molecular weight: 344mg

![Structure formula of the third chemical]
3- Test procedures

A series of different concentrations were prepared on the active ingredient basis (p.p.m) by diluting the formulated chemical in the solvent. About 5-8 concentrations, i.e. 25 -1600 ppm for each chemical of the three tested isolated chemicals were prepared. Thin film technique was used as a method of application, where the tested concentrations were applied through ethanol to the surface of a 0.9 cm diameter Petri – dish. One ml of each concentration of the tested plant chemicals was spread on the inner surface of the Petri dish, by moving the dish gently in circles. The Petri -dish used as control was treated with 1 ml ethanol only. The ethanol was evaporated under room temperature. Ten larvae of 4th or 2nd instar larvae of cotton leafworm were exposed about 24 hours in each per dish, then transferred to clean glass containers and fed on fresh castor leaves until the pupation period according to Ahmed(1985). Five replicates of each concentration were prepared. The larval mortality percentages of both 2nd and 4th instar were recorded and corrected according to the check by using Abbott’s formula, (1925). The data were then analyzed by using probit analysis (Finney,1971) and LC50 values were estimated for each tested chemical. The biological parameters of treated larval instars were evaluated as larval and pupal period, the pupation and adult emergence percentages, pupal weight. The fecundity, fertility, longevity and sex ratio of the adult stage treated as 4th instar larvae with LC50 values each of the phytochemicals were evaluated. Also, the observed malformations were recorded and photographed.

4- Separation technique.

The fresh plant(root and flower) was dried with out sunlight, after that was ground to fine powder. The ground material was transferred to a suitable container and soaked in methanol-methylene chloride (MeOH-CH2Cl2) (1:1) for 24 hrs at room temperature. The solvent (MeOH-CH2Cl2) was evaporated under reduced pressure to give a green oily residue. This oily material was quantitatively transferred by leading the material dissolved in the minimum amount of CH2Cl2.

On a small quantity of normal phase silica gel, then dried by evaporation the solvent under vacuum to the top of column chromatography, which packed with normal phase Silica gel (60, E- Merck) and eluted with hexane (3 L), followed by a gradient of hexane-CH2Cl2 up to 100% CH2Cl2 and CH2Cl2-MeOH up to 15% MeOH (2L of each solvent mixture). These fractions were further re-fractionated by column Sephadex LH-20 with eluted solvent of the following system: n-hexane, methylene chloride, methanol (7:4:0.5 or 7:4:1) and also by using HPLC (High Performance Liquid Chromatography)
Liquid Chromatography ) with eluted solvent of the following system: water, methanol (45:55 or 40:60) to give pure compounds in the end compounds in the end.

RESULTS AND DISCUSSION

1-Toxic effect

Data presented in Table(1) demonstrated that the three chemicals that isolated from three various plants, *Anaphalis margaritacea* L., *Chenopodium ambrosioides*, and *Ambrosia maritima* (L.) were effective against 2nd and 4th instar larvae of *S. littoralis*. The first chemical (*CdH$_2$O$_4$*) was the most toxic against 2nd and 4th instar larvae. The LC$_{50}$ values were 6 and 28 P.P.M., respectively. While the third chemical was the least toxic one as their LC$_{50}$ values were 60 and 84 P.P.M., respectively.

Table 1. Insecticidal activity of dibenzoc acid, chrysophan, and trimethoxy flavone against Spodoptera littoralis 2nd and 4th instar larvae.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>2nd instar</th>
<th>4th instar</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>LC$_{50}$</td>
<td>Slope</td>
</tr>
<tr>
<td></td>
<td>values</td>
<td>function</td>
</tr>
<tr>
<td></td>
<td>ppm</td>
<td></td>
</tr>
<tr>
<td><em>CdH$_2$O$_4$</em></td>
<td>6</td>
<td>6.1</td>
</tr>
<tr>
<td><em>CdH$_2$O$_4$</em></td>
<td>56</td>
<td>2.9</td>
</tr>
<tr>
<td><em>CdH$_2$O$_4$</em></td>
<td>60</td>
<td>6</td>
</tr>
</tbody>
</table>

These results agreement with those obtained by Mahmoud (2002) who demonstrated a toxic activity of four ethanolic extracts of *Cassia fistula*, *Ambrosia maritima*, *Tipuna tipu* and *Lantana camara* by using a contact method against 4th instar of *A. ipsilon* larvae. Also, Osman (1999) found that 40% of a chloroformic extract of *A. maritima* caused 100% mortality to *A. ipsilon* larvae, after 7 days of the treatment, using a contact method. Pizzale et al. (1997) proved that some isolated plant essential oils belonged to the family Asteraceae had insecticidal activity. These oils were analyzed by capillary gas chromatography-mass spectrometry(GC-MS) using two columns with different polarities(SE 52 and Carbowax). The antioxidant activity of these oils was evaluated with two methods: DPPH and RNA tests. Swidan (1994) showed a toxic activity of 25% mortality of *Ambrosia maritima*, against the 4th instar larvae of *S. littoralis*. Also, Abdallah and Kandil (1986) isolated and indicated the toxic activity of eleven fractions obtained from *Chenopodium ambrosioides*, *Banroof*, *Conya dicosroidis* and *Ulliga*, *Convulvulus arvensis*.
2. Latent effect

2.1. Larval and pupal periods

Data in Tables (2 and 3) showed that the second and fourth Spodoptera littoralis larvae treated at the LC50 values with the three tested chemicals increased the larval period. The effect was more pronounced with the first chemical (C10H18O4), where the larval period highly significant (p<0.01) increased to average 23 and 21 days for the larvae treated as 2nd and 4th instars, respectively, as compared to 14.5 and 11 days larval period of the check (untreated 2nd and 4th instar larvae, respectively). While the larval treatment of 2nd and 4th instars with the third chemical (C18H32O2) significantly (p<0.05) increased the larval period to average 20 and 16 days for the two instars, respectively. The second chemical (C6H12O4) caused none significant increase in the larval period to average 19.3 days for the two instars, respectively.

The larvae treated in the second instar larvae with the three tested chemicals at the LC50, highly significant (p<0.01) increased the pupal period of the resulting pupae to average 11.5, 9.4 and 9.5 days for the three chemicals, respectively (Table 2), as compared to 8.1 days for control. Whereas, the larvae treated in the fourth instar with the first chemical (C10H18O4) only highly significant (p<0.01) increased the pupal period to average 10.9 days, as compared to 7.1 days of the check (Table 3). While the larvae treated in the same instar with the second and third chemicals induced none significant increase for the pupal period to average 8 and 9.2 days for the two chemicals, respectively, as compared to 7.1 days of the check.

These results are in agreement to those obtained by Mahmoud (2002), who recorded an increase in the larval and pupal period of A. ipsilon as a result of the larval treatment of 4th instar with both C. fistula and A. maritima extracts by a contact method. Also, Osman (1999) found an increase in the larval and pupal period of A. ipselon after the larval treatment with a chloroformic extract of A. maritima using a contact method. Antonious et al. (1992) recorded that the longest pupal period was obtained when the larvae of S. littoralis treated with phytochemicals extracted by methanol, benzene or hexane of Dieffenbachia maculata and Adhatoda vasica.
Table 2. Effect of some phytochemicals on the larval and pupal development of *Spodoptera littoralis* treated as 2nd instar larvae with LC50 values.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Larval periods (days) ± SD</th>
<th>% Pupation ± SD</th>
<th>Pupal period (days) ± SD</th>
<th>Pupal weight (gm) ± S.E</th>
<th>% Moths emergence ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>C6H2O2</td>
<td>21.1±1.0**</td>
<td>51.8±1.6**</td>
<td>4.2</td>
<td>11.5±0.5**</td>
<td>0.216±0.06**</td>
</tr>
<tr>
<td>C6H5O2</td>
<td>19.3±0.2**</td>
<td>55.0±3.0**</td>
<td>11</td>
<td>9.4±0.31**</td>
<td>0.303±0.04**</td>
</tr>
<tr>
<td>C6H4O2</td>
<td>20.6±1.5**</td>
<td>99.7±1.6**</td>
<td>9</td>
<td>9.5±0.2**</td>
<td>0.266±0.01**</td>
</tr>
<tr>
<td>Check</td>
<td>14.5±0.6</td>
<td>100±0.0</td>
<td>0</td>
<td>8.1±0.4</td>
<td>0.365±0.05</td>
</tr>
<tr>
<td>F value</td>
<td>8.9</td>
<td>7.76.1</td>
<td>39.5</td>
<td>47.9</td>
<td>119.6</td>
</tr>
<tr>
<td>P value</td>
<td>0.01</td>
<td>0.003</td>
<td>0.002</td>
<td>0.001</td>
<td>0.0002</td>
</tr>
<tr>
<td>L.S.D at 5%</td>
<td>5.2</td>
<td>2.8</td>
<td>0.77</td>
<td>0.03</td>
<td>2.87</td>
</tr>
<tr>
<td>%</td>
<td>1</td>
<td>7.9</td>
<td>4.2</td>
<td>1.17</td>
<td>4.3</td>
</tr>
</tbody>
</table>

** = Highly Significant (p<0.01)
* = Significant (p<0.05)
S.D.= Standard deviation
Malfo.= Malformation%
L.S.D.= Least significant difference

Table 3. Effect of some phytochemicals on the larval and Pupal development of *Spodoptera littoralis* treated as 4th instar larvae with LC50 values.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Larval periods (days) ± SD</th>
<th>% Pupation ± SD</th>
<th>Pupal period (days) ± SD</th>
<th>Pupal weight (gm) ± S.E</th>
<th>% Moths emergence ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>C6H2O2</td>
<td>21.1±4.6**</td>
<td>55.0±5.0**</td>
<td>2.1</td>
<td>10.9±0.5**</td>
<td>0.244±0.05**</td>
</tr>
<tr>
<td>C6H5O2</td>
<td>15.0±1.0**</td>
<td>58.8±1.0**</td>
<td>7.2</td>
<td>9.0±1.8**</td>
<td>0.375±0.04**</td>
</tr>
<tr>
<td>C6H4O2</td>
<td>16.0±1.0**</td>
<td>65.6±2.1**</td>
<td>8</td>
<td>9.2±1.1**</td>
<td>0.329±0.01**</td>
</tr>
<tr>
<td>Check</td>
<td>11.0±2.0</td>
<td>100.0±0.0</td>
<td>0</td>
<td>7.1±0.8</td>
<td>0.440±0.05</td>
</tr>
<tr>
<td>F value</td>
<td>16.3</td>
<td>267.3</td>
<td>5.8</td>
<td>14.5</td>
<td>313.7</td>
</tr>
<tr>
<td>P value</td>
<td>0.003</td>
<td>0.00003</td>
<td>0.03</td>
<td>0.003</td>
<td>0.00005</td>
</tr>
<tr>
<td>L.S.D at 5%</td>
<td>4.3</td>
<td>4.4</td>
<td>2.4</td>
<td>0.06</td>
<td>3.7</td>
</tr>
<tr>
<td>%</td>
<td>1</td>
<td>6.1</td>
<td>6.5</td>
<td>3.6</td>
<td>5.6</td>
</tr>
</tbody>
</table>

** = Highly Significant (p<0.01)
* = Significant (p<0.05)
S.D.= Standard deviation
Malfo.= Malformation%
L.S.D.= Least significant difference
2.2. Pupation and Pupal weight

Tables (2 and 3) demonstrated that the larvae treated in 2\textsuperscript{nd} and 4\textsuperscript{th} instar with the three tested chemicals at the LC50, highly significantly (p<0.01) reduced the pupation period percent. The first chemical (C\textsubscript{3}H\textsubscript{6}O\textsubscript{3}) had the highest effect on pupation, it averaged 51.8 and 55 % for pupae treated as 2\textsuperscript{nd} and 4\textsuperscript{th} instar larvae, respectively, as compared with that of the check (100%). Whereas, the second (C\textsubscript{2}H\textsubscript{6}O\textsubscript{3}) and third (C\textsubscript{4}H\textsubscript{8}O\textsubscript{2}) chemicals had the next effect on the pupation, they averaged 55, 59.7 and 58.8, 65.6 % for pupae treated as 2\textsuperscript{nd} and 4\textsuperscript{th} instar larvae, respectively.

Likewise, the larval treatment of 2\textsuperscript{nd} and 4\textsuperscript{th} instar with the three tested chemicals at the LC50, highly significantly (p<0.01) reduced the pupal weight of the resulting pupae (Table 2 & 3). The first chemical was the most suppressive one on the pupal weight, it averaged 0.0216 and 0.0244 g for pupae treated as 2\textsuperscript{nd} and 4\textsuperscript{th} instar larvae, respectively, as compared to 0.365 and 0.440 g pupal weight of the check (pupae produced from untreated 2\textsuperscript{nd} and 4\textsuperscript{th} instar larvae, respectively). Whereas, the third (C\textsubscript{3}H\textsubscript{6}O\textsubscript{3}) and second (C\textsubscript{4}H\textsubscript{8}O\textsubscript{2}) chemicals induce the next decrease of the pupal weight, they averaged 0.286, 0.303 and 0.329, 0.375 g for pupae treated as 2\textsuperscript{nd} and 4\textsuperscript{th} instar larvae, respectively.

These results were similar to those obtained by Mahmoud (2002) who mentioned that the larval treatment of A. _ipsilon with C. fistula, A. maritima and T. tipu extracts highly significantly (p<0.01) reduced the pupation percent to about 50% of the control, and also decreased the weight of the resulting pupae. Also, Osman (1999) demonstrated a decrease in the pupal weight of A. _ipsilon following treatment of larvae with a chloroformic extract of A. maritima using a contact method Amr et al. (1995) indicated a reduced pupal weight of S. littoralis treated as larvae with a chloroformic and ethanolic extracts of Nerium oleander leaves. There are similar results were obtained by Abo-El-Ghazal et al. (1994) who showed a significant reduction in the pupal weight of A. _ipsilon treated as larvae with acetone and ethanolic extracts of Vinca rosea and Melia azedarach and also they added that the larval feeding on the ethanolic extracts at concentrations of 0.02 and 0.01% completely failed to pupate. Also, Antonious (1992) recorded that the castor oil leaves treated with Dieffenbachia maculata extracts proved to be more effective than those treated with Adhatoda vasica extracts in reducing the pupation of S. littoralis.
2.3. Moths emergence

Data in Tables (2 & 3) showed that second and fourth instar larvae of *S. littoralis* treated with the three tested chemicals at the LC50 values induce highly significant (p<0.01) reduction in the moths emergence. The first chemical (C6H14O4) had the highest effect on the moths emergence, where it averaged 30.6 and 55.7 % for adults treated as 2nd and 4th instar larvae respectively, as compared to 57.4 and 58.9 % adult emergence of the check (adults produced from untreated 2nd and 4th instar larvae, respectively). While the larval treatment of the two instars with the third chemical (C6H12O4) had the next effect on the adult emergence to average 51 and 64 % for adults treated as 2nd and 4th instar larvae, respectively. Whereas, the second (C9H18O4) chemical had the least effect on the adults emergence to average 67 and 71% for adults treated as 2rd and 4th instar larvae, respectively.

These results are agreement with those obtained by Mahmoud (2002) who demonstrated that the larval treatment of 4th instar of *A. ipsilon* with *A. maritima* extract induced the highest reduction in the adult emergence by a contact method. Also, Abo-El-Ghar et al. (1994) recorded a decrease in the adult emergence of *A. ipsilon* treated as larvae instar with petroleum ether extracts of *L. cylindrica*, *A. majus*, *C. elegans* and *B. rosea*, as compared to control. Antonious et al. (1992) found that a decrease in the adult emergence of *S. littoralis* after the larval treatment with ethanolic or methanolic extracts of *D. maculata* and *A. vasica* plants.

Table 4. Effect of some phytochemicals on the adults of *Spodoptera littoralis* treated as 4th instar larvae with LC50 values.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Fecundity eggs/ f</th>
<th>Fertility eggs/ f</th>
<th>Longevity (days)</th>
<th>Adult sex ratio(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± S.D.</td>
<td>Mean ± S.D.</td>
<td>Mean ± S.D.</td>
<td>Male %</td>
</tr>
<tr>
<td>C6H14O4</td>
<td>93.3 ± 1.5&quot;</td>
<td>89.2 ± 1.3&quot;</td>
<td>8.0 ± 1.3&quot;</td>
<td>44.4%</td>
</tr>
<tr>
<td>C9H18O4</td>
<td>110.8 ± 5.0&quot;</td>
<td>104.6 ± 1.95&quot;</td>
<td>12.5 ± 1.5&quot;</td>
<td>40.0%</td>
</tr>
<tr>
<td>C6H12O4</td>
<td>115.8 ± 0.8&quot;</td>
<td>109.0 ± 2.7&quot;</td>
<td>8.5 ± 1.0&quot;</td>
<td>43.3%</td>
</tr>
<tr>
<td>Check</td>
<td>568.7 ± 1.66</td>
<td>569.6 ± 8.6</td>
<td>15.2 ± 0.7</td>
<td>39.5%</td>
</tr>
<tr>
<td>F value</td>
<td>4394.16</td>
<td>4462.6</td>
<td>21.7</td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>0.000004</td>
<td>0.000001</td>
<td>0.001</td>
<td></td>
</tr>
<tr>
<td>LSD at 5%</td>
<td>3.7</td>
<td>10.5</td>
<td>2.6</td>
<td></td>
</tr>
<tr>
<td>LSD at 1%</td>
<td>5.7</td>
<td>16.3</td>
<td>3.9</td>
<td></td>
</tr>
</tbody>
</table>

** = Highly Significant (p<0.01)  * = Significant (p<0.05)
2.4. Adult fecundity and fertility

Data presented in Table(4) showed that the treatment of 4th instar larvae of S. littoralis with the three tested chemicals at the LC50 values highly significant (p<0.01) retard the fecundity of the emerged adults. The first chemical (C2H2O2) had the strongest effect on the fecundity, where it reduced the fecundity to average 93.3 eggs/f, as compared to 558.7 eggs/f of the check. While the third (C3H6O2) and second (C2H2O2) chemicals had the next effect on the adult fecundity, where they reduced the adult fecundity to average 115.8 and 110.8 eggs/f for adults treated as 4th instar with the two chemicals, respectively.

Likewise, the larval treatment of 4th instar of S. littoralis with the three tested chemicals at the LC50 values highly significant (p<0.01) inhibited the eggs fertility (Table,4). The first chemical (C1H1O4) had the highest effect on eggs fertility, where it reduced the eggs fertility to 85.2 eggs/f, as compared to 509.6 eggs/f of the check. Whereas, the third (C3H6O2) and second (C2H2O2) chemicals had the following effect on the eggs fertility, it averaged 109 and 104.6 eggs/f for adults treated as 4th instar larvae with the two compounds, respectively.

These results were similar to those obtained by Mahmoud (2002) who recorded that a reduction in the adult fecundity and fertility of A. ipsilon treated as 4th instar larvae with both A. maritima and C. fistula extracts by a contact method. Also, Osman (1999) cleared that a decrease in the percent of eggs hatching of A. ipsilon treated with chloroform extracts of A. maritima, via oral administration. Whereas, Abo-El – Ghar et al. (1994) showed fecundity deterrent effects of plant extracts against A. ipsilon were found in the polar (acetone, ethanol and water) extracts of M. azedarach, V. rosea, L.cylindrica, P. vera, C. elegans, A. majus and A. graveolens rather than non polar (petroleum ether).

2.5. Adult longevity

Table(4) indicated that the 4th instar larvae of S. littoralis treated with the three tested chemicals at the LC50 values caused a highly significant (p<0.01) decrease in the longevity of the emerged adults, as compared to control. The first chemical (C2H2O2) had the highest effect in reducing of adult longevity to average 8 days, as compared to 15 days of control. Whereas the third chemical had the next effect on the adult longevity, where it averaged 8.5 days. While the least shorten of the longevity was obtained by the larval treatment of 4th instar larvae with the second chemical (C2H2O2), it averaged 12.6 days.
These results agreement with that obtained by Mahmoud (2002) who recorded a significant decrease in the adult longevity of *A. ipsilon* by the larval treatment of 4th instar with *A. maritima* and *T. tipu* extracts by a contact method. Osman (1999) mentioned that chloroform extract of *A. maritima* decreased the adult longevity of *A. ipsilon* treated as larvae with LC50 values, using a contact method.

2.6. Adult sex ratio

Data obtained in Table (4) showed that the larval treatment of 4th instar of *S. littoralis* with the three tested chemicals at the LC50 values shifted the sex ratio, it increased the males and decreased the females than that of the check. The effect was more pronounced with the first and third chemicals, where it reached 44.4, 43.3, and 55.6, 56.7% for both adult males and females respectively, as compared to 39.5 and 60.5% of adult males and females respectively, of the check. While the sex ratio of both males and females did not differ than that of the check at the larval treatment of 4th instar with the second compound (C27 H30 O4), it reached to 40 and 60% for both adult males and females respectively, treated as 4th instar.

2.7. Morphogenetic effects

Data presented in Tables (2 & 3) showed that the larval treatment of 2nd and 4th instars of *S. littoralis* with the three tested chemicals at the LC50 values induced a noticeable increase in the pupal malformations, as compared to the check. The second (C27 H30 O4) and third (C28 H52 O7) chemicals induce the highest percent, it reached 11, 9 and 7.2, 8% for pupae treated as 2nd and 4th instar larvae with the two chemicals, respectively, as compared to 0% pupal malformations of the check. Whereas, the first chemical (C21 H34 O4) recorded the lowest percent, it reached 4.2 and 2.1% for pupae treated as 2nd and 4th instar larvae, respectively.

With regarded to the adult malformations (Tables 2 & 3), it was found that the larval treatment of 2nd and 4th instar of *S. littoralis* with the first (C21 H34 O4) and third (C28 H52 O7) chemicals at the LC50 values caused the highest percent of adult malformations, it reached 3.7, 3.5 and 4.1, 3.6% for adults treated as 2nd and 4th instar larvae with the two chemicals, respectively, as compared to 0% of the check. Whereas, the second chemical (C27 H30 O4) induce the next percent of adult malformations, it reached 3.4 and 2.9% for adults treated as 2nd and 4th instars, respectively, as compared to control (0%).

These results are similar to that obtained by Mahmoud (2002), who demonstrated that the pupal and adult malformation percent as result of the larval treatment of 4th instar larvae of *A. ipsilon* with the *A. maritima*, *T. tipu*, *L. camara*
and *C.fistula* extracts, the effect was more pronounced with both *A. maritima* and *T. tipu* extracts, where the highest pupal and adult malformations were recorded by the larval treatment with the two extracts, while the other *C.fistula* and *L. camara* extracts had the least effect one. Also, the same results obtained by Abo-El-Ghar et al. (1994) on *A. ipsilon* and Antionious et al. (1992) on *S. littoralis*.

Malformations of *S. littoralis* pupae resulting from the larval treatment of 2nd and 4th instars in the present work mostly appeared as malformed prepupa failed to cast the old cuticle with complete blackening of the body leading to death (Fig.1), or abnormal pupae showing body shrinkage or undersized pupae (Fig.2) or malformed pupa failed to cast the larval skin (Fig.3). Adult malformations often appeared as poorly developed moths failed to emerge from the pupal cast (Fig.4) or a moth showing poorly developed and deformed twisted wings (Fig.5 and 6), as compared to the pupa and adult of the control (Fig.7 and 8).
Fig 1. Malformation prepupa failed to cast the old cuticle with complete blackening of the body leading to death

Fig 2. Abnormal pupa showing body shrinkage or undersized pupa

Fig 3. Malformation pupa failed to cast the larval skin

Fig 4. Adult malformations often appeared as poorly moth failed to emerge from the pupal cast

Fig 5. and 6. Moth showing poorly developed and deformed twisted wings

Fig 7. and 8. Normal pupa and adults
REFERENCES


active compounds from extracts of *Mimtea barnroof* and Ullaiaq. Bull. 

extracts on development and fecundity of *Agrotis ipsilon*. Bull. Ent. Soc. 
Egypt, 21:171-190.


leaves extract on some physiological changes of the cotton leafworm, 

of feeding the cotton leafworm, *Spodoptera littoralis* on certain plant leaves, 3: 
Latent effect of larval feeding. International symposium crop protection, 
Mediterranean-South Agricultural Research and Development Project, 
Rijksuniversiteit Gent, 57:707-713.

studies on the Egyptian cotton leafworm, *Prodenia litura*: Susceptibility of 
different larval instars of *Prodenia* to insecticides. J. Econ. Entomol., 57:591-59.


some plant extracts on the black cutworm, *Agrotis ipsilon*. Ph. D. Thesis, Ain 
shams Univ. Cairo. Rijksuniversiteit Gent, 57:707-713.

Chemical composition and antioxidant activity of essential oils of different botanical 
Origin. Dipartimento di Scienze degli Alimenti, Universita degli Studi di Udine, Via 
Marangoni, 33100 Udine, Italy.

and their common constituent on the psyllid pest, *Heteropsylla cubana* of social 


تأثير السمى والمتاثر لثلاث مركبات كيميائية مأخوذة على برقتان

دودة ورق القطن الكبرى

الهام فاروق محمود عبد الراحم، عماد محمد أحمد
أحمد عبد الله أحمد، طارق عبد الحليم محمد

1. مختبر بحوث سدود، معهد وآلاف الديانات، مركز البحث الزراعي، كلية العلوم، قسم الكيمياء، جامعة المنيا.
2. إجريت هذه الدراسة بغرض تقييم تأثير السمى لثلاث مركبات معروفة من نباتات الأزهار والزهيدية والزهيدية والزهيدية. اختُبرت عينات من برقتان عمراني وبرقتان دورات ورق القطن الكبير. كانت الصيغة الجزيئية للمركبات المختبرات هي: 

\[ \text{C}_{6}H_{12}O_{6}, \text{C}_{6}H_{12}O_{6} \text{H}_{2} \text{SO}_{4}, \text{C}_{6}H_{12}O_{6} \text{H}_{2} \text{SO}_{4} \]

تركزت برقات عمران ما بين 24 ساعة مع فتم التعرض بكميات مختلفة لكل مركب في درجة الماء. وقد وجدت أن المركب الأول كان له العنصر الأول والثاني، بينما كان المركب الثاني له العنصر الثاني، حيث بلغت قيمة تركيز القصفي له (60.84 ppm) على كل هذين المركبين على الترتيب. تأثرت المعايير البيولوجية للبرقات المختبرات، وقد كان التأثير كان أكثر وضوحًا مع معدل معدل برقات عمران والثاني والرابع بمرور الوقت حيث أدى ذلك إلى التطور في العبع البراعم والثدي والخلي ونسج من نسب التطور والعدو والدواء، ودرجة الصعوبة مما أدى إلى خسر في عمر البراعم الخفيفة و wnętrرح في أنماط الجسيم لل nokwالماء مثلا: 

للمراكب الثلاثة حفرات العين المعاملة. كما أجريت المحاكمة الشاملة برقة العمر الثاني والرابع بالمركبات الثلاثة نسب من المركبات في مجموع العير والريع للبرقات المختبرة. وقد لوحظ في بعض الأنشطة البيولوجية المختلفة للبرقات المختبرة بالمركبات الثلاثة إن الرعاية بالمركبات الثالثة أخذت تأثيرًا إفراز النتائج التالي: المركب الأول مقارنة بعمليات المركب الثاني.