CALCIUM PHOSPHATE AS AN ADJUVANT FOR INACTIVATED RIFT VALLEY FEVER VACCINE

ELLIAN, KHAIRAT A., LILLY S. SALAMA, A. M. IBRAHIM, TARADI ABD - EL FATTEH AND A. M. DAOUD

Veterinary Serum and Vaccine Research Institute, ARC, Egypt

(Manuscript received 21 February 2005)

Abstract

Vaccination to protect animal against infectious diseases may be enhanced by using adjuvants that can selectively stimulate immunoregulatory responses. A novel adjuvant composed of calcium phosphate (CAP) was compared with the commonly used aluminium hydroxide (alum) gel adjuvant for its ability to induce immunity to Rift Valley Fever virus. Results indicated that CAP was more potent as an adjuvant than alum, induced high titres of antibody. Furthermore, it is economical, simple to manufacture and it is a natural constituent of the animal body.

INTRODUCTION

Rift Valley Fever (RVF) is an arthropod-borne viral disease affecting animals and human. It is an economically important viral disease and widely distributed in different localities of Africa and Asia where periodic epizoqtic and epidemic occurred causing heavy losses among lambs and calves (Woods *et al.*, 2002 and Fagbo, 2002).

RVF disease is caused by RNA single stranded virus belonging to family Bunyaviridae (WHO, 1982 and Connie, 1996). The appearance of RVF disease in Egypt in 1977 (Imam *et al.*, 1977), and its reappearance in 1993 (El-Gabery *et al.*, 1994) increased the demand to develop a potent inactivated RVF vaccine. The adjuvants are modulators of the immune system and their function is to improve vaccine efficacy in order to offer protection against infection. One of these adjuvants is calcium phosphate which is a novel adjuvant elicited little or no inflammation at the site of inoculation and induced higher titre of neutralizing antibody (Bio Sante Pharmaceuticals Inc., 2000). Alum compounds are the most extensively used adjuvants in licensed vaccines. Although they effectively enhance immune response, they cause severe inflammatory reaction at the injection site and the duration of this inflammation is somewhat long as cited by Goto *et al.* (1997) who reported that the local tissue reactions caused by injection of CAP gel completely ceased by the 4th week, while irritation caused by aluminium hydroxide gel persisted for 8 weeks.

The aim of this work is to study the effect of CAP (as an adjuvant) when added to Rift Valley Fever binary inactivated virus on the immune response of vaccinated sheep.

MATERIALS AND METHODS

1. Animals

1.1. Mice (Swiss albino mice)

1.1.1. Adult mice

Twenty one (twenty eight - day old) mice were used for toxicity and potency test for both calcium phosphate and vaccines, respectively.

1.1.2. Baby mice

Three (five-day old) mice were used for safety of the prepared inactivated virus.

1.2. Sheep

- **1.2.1.** Fourteen susceptible balady sheep six month age were used for the potency test of the vaccines.
- **1.2.2.** Eight lambs (5-10- day old) were used for safety of the RVF vaccine with different calcium phosphate concentrations.

2. Virus

RVF virus ZH-501 with a titre of 7.5 \log_{10} TCID_{S0}/ml were kindly supplied by RVF Department, Veterinary Serum and Vaccine Res. Inst., Abbasia, Cairo. It was isolated from human patient in Zagazig, Sharquia Province during outbreak in 1977.

3. Conjugate

Horseradish peroxidase conjugate labeled antispecies (antisheep) was purchased from Sigma Company. It was diluted in PBS immediately before use for ELISA test.

4. Adjuvant

1.1.1. Aluminium hydroxide gel

2 % gel was purchased from Honil Limited, London, United Kingdom.

1.2.1. Calcium phosphate (CAP)

Composed of:

- 1. Calcium chloride (Winlab).
- 2. Dibasic sodium phosphate (El-Nasr Pharmaceutical Chemicals Co.).
- 3. Sodium citrate (Analar).

It was prepared according to Bio Sante Pharmaceuticals, Inc., Smyrna, Georgia (2000).

Toxicity test

Adult mice were used for the toxicity of CAP adjuvant in vaccine preparation. Three groups of mice (15 per each), one inoculated I/P and the second S/C while, the third group was kept as a control, and all groups were observed for 10 days post-inoculation.

Preparation of the vaccine

1. Virus

RVF ZH-501 was inactivated by binary ethyleneimine according to M, Eman (1995), then different forms of vaccines were prepared, one with 25 % aluminium hydroxide gel and the three others with 75 %, 50 % and 25 % calcium phosphate, respectively.

2. Addition of CAP adjuvant

CAP was added with different concentrations to the inactivated virus as 75 %, 50 % and 25 %.

Evaluation of the vaccine

1. Sterility test

It was done according to OIE (2000).

2. Safety test

It was performed according to El-Nimr (1980) and M, Eman (1995).

a. Baby mice

They were inoculated I/C.

b. Lambs

Nine lambs were inoculated each with 10 ml of the vaccine (5 ml I/P and 5 ml S/C), then these animals were observed for 10 days for any sign of RVF disease or death (El-Nimr, 1980 and Eman, 1995).

Potency test

Adult mice were inoculated I/P by two doses of the vaccine one week apart, and then challenged to calculate the ED₅₀ for each formula of the vaccine separately according to Randall *et al.* (1964).

Experimental Design

Fourteen balady sheep were divided into 5 groups:

- Group (1): Three sheep were vaccinated S/C with inactivated RVF vaccine with 75 %
- **Group (2):** Three sheep were vaccinated S/C with inactivated RVF vaccine with 50 % CAP.
- **Group (3):** Three sheep were vaccinated S/C with inactivated RVF vaccine with 25 % CAP.
- Group (4): Three sheep were vaccinated S/C with inactivated RVF vaccine with

aluminium hydroxide gel (commercial one).

Group (5): Two sheep were kept as control (non-vaccinated).

All animals were observed for 6 months post- inoculation for sero-conversion.

Serological tests

1. Serum neutralization test

It was done according to Walker (1975).

2. Indirect enzyme linked immunosorbent assay (indirect ELISA)

It was done according to Voller et al. (1976).

RESULTS AND DISCUSSION

Table 1. Results of toxicity test in mice.

Adjuvant	Mice	- Control		
	s/c	I/P	Control	
CAP	0/15*	0/15*	0/15*	

^{*} Number of dead mice over number of survived mice.

Table 2. Results of sterility, safety and potency test of the prepared vaccine.

		Safe	Potency*	
Type of vaccine	Sterility	Baby mice**	Lamb***	ED ₅₀ /ml
CAP 75 %	Sterile	0/8	0/2	0.0006/ml
CAP 50 %	Sterile	0/8	0/2	0.0003/ml
CAP 25 %	Sterile	0/8	0/2	0.0005/ml
luminium hydroxide gel	Sterile	0/8	0/2	0.0006/ml

^{*}The minimum permissible limit of ED $_{50}$ /ml is 0.02 ml.

^{**} Safety test in baby mice performed of inactivated RVF virus without adjuvant.

^{***} Safety in lamb = No thermal or dinical reaction or manifestation.

Table 3. Result of neutralizing antibody index (NI) of sheep sera vaccinated with different formula of RVF prepared vaccines.

of .	Types of	NO. of	Before Vaccination	Neutralizing Indices Weeks post Vaccination								
	different											
	adjuvant			1	2	3	4	8	12	16	20	24
G1	CAP 75	3	0.4	1.0	1.4	1.7	2.4	2.7	3.0	2.6	2.0	1.5
			0.3	0.7	1.0	1.4	1.7	2.0	2.4	2.5	2.1	1.7
	%*		0.3	0.7	1.0	1.7	2.0	2.4	2.7	2.5	2.5	1.8
		Mean	0.33	0.8	1.1	1.6	2.0	2.4	2.7	2.5	2.2	1.6
G2	-	3	0.4	0.7	1.4	1.7	2.4	2.7	3.0	3.3	3.7	3.4
			0.3	1.4	1.7	2.0	2.7	3.0	3.4	3.7	4.0	4.0
	CAP50%*		0.4	1.0	1.4	2.0	2.4	3.0	3.2	3.4	3.7	3.7
		Mean	0.36	1.0	1.5	1.9	2.5	2.9	3.2	3.5	3.8	3.7
G3	CAP 25	3	0.4	0.7	1.4	1.7	2.0	2.4	2.5	3.0	2.7	2.5
			0.4	0.7	1.4	1.7	2.0	2.4	2.7	3.7	3.0	2.9
	%*		0.3	1.0	1.0	1.7	2.4	2.7	3.0	3.7	3.4	2.9
	-	Mean	0.36	0.8	1.2	1.7	2.13	2.5	2.7	3.4	3.0	2.8
G4	Alum gel	Alum gel	0.4	0.7	1.4	1.7	2.0	2.4	2.4	2.0	1.7	1.4
			0.3	1.0	1.0	1.4	1.7	2.0	2.7	2.7	2.0	1.
			0.4	0.7	1.4	1.7	2.0	2.7	2.7	2.7	2.4	1.
		Mean	0.36	0.8	1.2	1.6	2.0	2.3	2.6	2.3	2.0	1.
GS .	control	2	0.3	0.2	0.4	0.3	0.3	0.2	0.3	0.3	0.2	0.
			0.1	0.3	0.2	0.3	0.4	0.2	0.2	0.3	0.3	0.
		Mean	0.2	0.25	0.3	0.3	0.35	0.2	0.25	0.3	0.25	0.

G1: Binary inactivated RVF vaccine with 75 % of CAP

G2: Binary inactivated RVF vaccine with 50% of CAP

G3: Binary inactivated RVF vaccine with 25 % of CAP

G4: Binary inactivated RVF vaccine with alum hydroxide gel

G5: Control (non-vaccinated).

^{*} CAP: Calcium phosphate

CALCIUM PHOSPHATE AS AN ADJUVANT FOR INACTIVATED RIFT VALLEY FEVER VACCINE

Table 4. Result of indirect Elisa technique of sheep sera vaccinated with different formula of RVF prepared vaccines

Group of animals*	Types different adjuvant**	NO. of animals	Before Vaccination	Optical Density Weeks post Vaccination								
				G1		3	0.020	0.057	0.062	0.074	0.088	0.096
*CAP	0.010	0.051	0.059		0.062		0.076	0.088	0.103	0.094	0.082	0.0
75 %	0.012	0.044	0.060		0.069		0.080	0.090	0.114	0.101	0.098	0.0
	Mean	0.014	0.050		0.060	0.068	0.081	0.091	0.109	0.098	0.091	0.0
		. 3	0.011	0.055	0.065	0.079	0.084	0.096	0.110	0.134	0.145	0.1
G2	CAP		0.020	0.062	0.069	0.082	0.096	0.102	0.131	0.141	0.158	0.1
	50 %*		0.017	0.059	0.062	0.078	0.089	0.102	0.127	0.139	0.150	0.1
		Mean	0.016	0.058	0.065	0.079	0.089	0.100	0.122	0.138	0.151	0.1
			0.011	0.056	0.061	0.076	0.086	0.097	0.117	0.122	0.117	0.1
G3	CAP		0.021	0.055	0.063	0.069	0.081	0.091	0.112	0.128	0.125	0.1
63	25 %*		0.17	0.061	0.064	0.071	0.079	0.101	0.113	0.125	0.119	0.1
		Mean	0.016	0.057	0.062	0.072	0.082	0.096	0.114	0.125	0.120	0.1
G4	Alum gel	gel 3	0.021	0.055	0.057	0.067	0.074	0.089	- 0.104	0.095	0.089	0.0
			0.013	0.066	0.059	0.069	0.071	0.086	0.111	0.087	0.077	0.0
			0.014	0.051	0.061	0.071	0.078	0.090	0.107	0.090	0.081	0.0
		Mean	0.016	0.057	0.059	0.069	0.074	0.088	0.107	0.090	0.082	0.0
G5	control	2 -	0.011	0.012	0.010	0.018	0.019	0.013	0.020	0.012	0.020	0.0
			0.020	0.017	0.021	0.019	0.022	0.017	0.011	0.010	0.011	0.0
		Mean	0.015	0.014	0.015	0.018	0.020	0.015	0.015	0.011	0.015	0.0

G1: Binary inactivated RVF vaccine with 75 % of CAP

G2: Binary inactivated RVF vaccine with 50 % of CAP

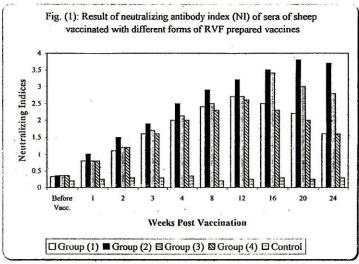
G3: Binary inactivated RVF vaccine with 25 % of CAP

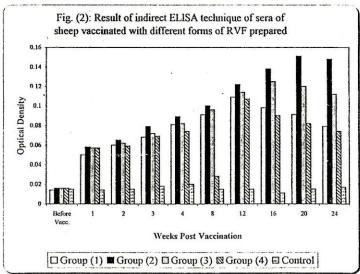
G4: Binary inactivated RVF vaccine with alum hydroxide gel

G5: Control non-vaccinated

* CAP: Calcium phosphate

- cut - off value= 0.03





When the toxicity test was carried out on adult mice, the result revealed that neither S/C nor I/P routes of injection elicited inflammation at the site of injection or any sign of toxicity during the test as shown in Table 1. All batches of the prepared vaccines were sterile and safe when inoculated in baby mice and lambs which showed no variation of body temperature of lambs or no signs of illness and no deaths were observed in mice and lambs. The most potent vaccine was that containing 50 % calcium phosphate as an adjuvant as its ED $_{50}$ /ml was 0.0003/ml followed by vaccine containing 25 % CAP as an adjuvant as its ED $_{50}$ /ml was 0.0005/ml and the last one was 75 % calcium phosphate as its ED $_{50}$ /ml was 0.0006/ml as shown in Table 2. All these batches were within the permissible limit as cited by Randall *et al.* (1964) who said that the ED $_{50}$ must not be more than 0.02/ml. The ED $_{50}$ of aluminium hydroxide gel vaccine batch was 0.0006/ml. These results agreed with K, Gehan (1990) and M, Eman (1995), when they used alum gel vaccine as there is no available data on RVF vaccine adjuvated with calcium phosphate.

The immune response of vaccinated sheep was tested by SNT (Table 3 and Figure 1) and showed the neutralizing indices of all groups of sheep. It was noticed and sera of sheep vaccinated with RVF vaccine with 50 % calcium phosphate gave the highest level at second week (mean of NI equal 1.5) as Pini *et al.* (1973) suggested that the protective titre was 1.5 log₁₀ TCID₅₀/ml and reached its peak at 20th week post vaccination with a mean of NI 3.8. This agreed with Biosante Pharmaceuticals Inc. (2000) who found that Herp's simplex virus type two (HSV-2) plus calcium phosphate gave high antibody level at 6 weeks after immunization and still was high up to the week fourteen. In case of RVF vaccine with 25 % CAP adjuvant, the protective level appeared at 3rd week post-vaccination with an average of 1.7 NI, while RVF vaccine with 75 % adjuvant, the protective level appeared at post vaccination with an average of 1.6 NI and reached its peak at the 16th week post vaccination being 3.4 NI. The protective level of the vaccine of aluminium hydroxide gel appeared at 3rd week post vaccination with an average 1.6 NI and reached its peak at 12th week post vaccination being 2.6 NI.

The result of SNT was correlated with that obtained by ELISA test as shown in Table 4 and Figure 2. This agreed with M, Eman (1995) and Hassan *et al.* (2001), but they used inactivated Rift Valley Fever inactivated aluminium hydroxide gel vaccine.

From the previous data, CAP adjuvant induces no inflammation at site of entry and induces immunological enhancement without toxicity and gives a higher titer of antibody earlier than aluminium hydroxide gel, furthermore, it is easy to manufacture on an industrial scale and shows insignificant variation in quality and physicochemical properties between batches in production condition.

REFERENCES

- Bio Sante Pharmaceuticals, Inc., Smyrna, Georgia. 2000. Calcium phosphate nanoparticle adjuvant. Clinical and Diagnostic Laboratory Immunology, November, 2000 Vol. 7, No. 6: 899 - 903.
- Connie, S. S. 1996. Bunyaviridae: The viruses and their replication. Field Virology.
 3rd ed. Vol. 1, Chapter 47. Philadelphia, Lippincott, Raven.
- El-Gabery, G. H., Nawal M. A., Hadia. A., Fathia M. M. and N. N. Ayoub. 1994.
 Unclassical picture of RVF in man and animals in Aswan governorate in May 1993.
 Vet. Med. J., Giza, 42 (1): 135-138.
- El-Nimr, M. M. 1980. Studies on the inactivated vaccine against RVF. Thesis, Ph.D. Fac. Vet. Med., Assiut Univ., Egypt.
- Eman. M. S. 1995. Studies on Rift Valley Fever vaccine inactivated with binary. Ph.
 D. thesis Vet. Sc., Fac. Vet. Med., Cairo Univ., Egypt.
- Fagbo, S. F. 2002. The involving transmission pattern of Rift Valley Fever in the Arabian Peninsula. AMNY Acad. Sci., 2002 Oct., 969: 201 - 204.
- Gehan, K. M. 1990. Studies on Rift Valley Fever among animals in Egypt. Ph. D. Thesis, Fac. Vet. Med., Zagazig Univ., Egypt.
- Goto, N., H. Kato and J. I. Maeyama. 1997. Local tissue irritating effects and adjuvant activities of calcium phosphate and aluminium hydroxide with different physical properties. Vaccine, 15: 1364 - 1370.
- Hassan, K. Z., K. A. Elian and M. M. Taha. 2001. Some studies on sheep vaccinated with Smithburn attenuated Rift Valley Fever vaccine. Egypt. J. Agric. Res., 79 (3).
- Imam, Z. E. I., M. A. A. Darwish, R. El-Karamany and F. Omar. 1977. A preliminary report on an epidemic of Rift Valley Fever in Egypt. J. Egyptian Public Health. Assoc., 52: 417 - 418.
- 11. OIE. 2000. Rift Valley Fever.
- Pini, A., L. J. Lund and S. J. Davis. 1973. Fluorescent and neutralizing antibody response to injection by RVF. J. S. Afr. Z. Med. Ass., 44 (11): 161 - 165.
- Randall, R., L. N. Binn and V. R. Harrison. 1964. Immunization against Rift Valley
 Fever virus, studies on the immunogenicity of lyophilized formalin inactivated
 vaccine. J. Immun., 93 (2): 293 299.
- 14. Voller, A., D. E. Bidwell and A. Bartlett. 1976. Enzyme immunoassay in diagnostic

- Voller, A., D. E. Bidwell and A. Bartlett. 1976. Enzyme immunoassay in diagnostic medicine, theory and practice. Bull. W. H. O., 53: 55 - 56.
- Walker, S. J. 1975. Rift Valley Fever: A review committee on forgein animal diseases, United State of Animal Health Association, US Army Med., Bes. Inst. Inf. Dis., Fredrick, Maryland.
- WHO. 1982. Rift Valley Fever and emergan human and animal problem. WHO Offest, Publication No. 63.
- Woods, C. W., A. M. Karpati, T. Greint, W. McCarthy, P. Galuruku, E. Muchiri, L. Dunster, A. Henderson, A. S. Khan, R. Swanepoel, I. Bonmarin, R. Marlin, P. Mann, B. L. Smoak, M. Ryan, T. G. Ksiazek, R. K. Arthur, N. D. Kuyeze, N. N. Agafa, C. Peters and WHO. 2002. Hemorrhagic fever task force. Emerging infectious Diseases, 8 (2).

إستخدام فوسفات الكالسيوم كمحسن مناعى للقاح حمى الوادى المتصدع المثبط

خيرات عبد المجيد عليان ، للى صبحى سلامة ، ألفونس مينا إبراهيم ، تراضى عبد الفتاح ، أحمد محمود داود

معهد بحوث الأمصال واللقاحات البيطرية – مركز البحوث الزراعية- وزارة الزراعة- النقى-جيزة – مصر

إن التحصين هو الوسيلة لوقاية الحيوانات ضد الأمراض الوبائية ويمكن رفع المستوى المناعى للحيوانات بإضافة بعض المحسنات للقاحات. وقد استخدم محسن جديد في هذا البحث وهـو فوسـفات الكالسيوم وبمقارنته بالالومنيوم هيدروكسيد جل في هذا البحث وجد أنه أقوى ويعطى أجسـامًا مناعيـة بنسبة أعلى من الالومنيوم هيدروكسيد جل. بالإضافة إلى أنه مادة أقتصادية وسهلة التصنيع كمـا أنهـا أحدى المكونات الطبيعية في جسم الحيوان.